



20/21/22 MARZO 2013  
PADENGHE SUL GARDA - BS



**Workshop Alifax**  
**TECNOLOGIA LIGHT SCATTERING: NUOVO APPROCCIO**  
**DIAGNOSTICO AL PAZIENTE CRITICO**  
**Diagnostica batteriologica rapida dei liquidi biologici**  
**e CVC**

*Carla Fontana*





## L'ossessione della rapidità

- La vera sfida per il microbiologo del futuro, non è più solo l'accuratezza e l'appropriatezza, ma arrivare in tempo
- Perchè produrre un referto impeccabile, ma in ritardo serve solo per la storia, non aiuta il clinico e non serve al paziente
- Tutta la spinta e l'innovazione degli ultimissimi anni è stata verso la contrazione dei tempi
- Il TAT è divenuto la nostra guida



## Agire sul TAT

Il TAT è una variabile dipendente ed esistono due accezioni del concetto di TAT:

- Il TAT “**clinico**” calcolato dal prelievo al referto, dal prelievo all’outcome (Misura del processo nel suo insieme)
- Il TAT “**metodologico**”, o di metodo ossia il tempo della procedura scelta per arrivare a finalizzare un referto (misura delle scelte operate dal microbiologo)
- Su entrambi pesa in maniera significativa la fase preanalitica



## Preanalitica

- Il grande balzo in termini di rapidità, ma anche di efficienza è stato poter agire sulla fase preanalitica sotto due aspetti
  - il primo: lavorare i liquidi biologici in mezzo liquido, riducendo all'essenziale la coltura su piastra (notoriamente lenta e poco efficiente) e colmando i danni del trasporto/conservazione del campione
  - il secondo portare tutti o pressocchè tutti i campioni microbiologici in fase liquida



## **La storia di questo cambiamento**



# La nostra esperienza

Received: 2007.08.14  
Accepted: 2008.06.03  
Published: 2009.02.01

## A novel culturing system for fluid samples

### Authors' Contribution:

- A** Study Design
- B** Data Collection
- C** Statistical Analysis
- D** Data Interpretation
- E** Manuscript Preparation
- F** Literature Search
- G** Funds Collection

Carla Fontana<sup>1,2A,B,C,D,E,F,G</sup>, Marco Favaro<sup>1B,C,D,F</sup>, Silvia Minelli<sup>2B,D</sup>, Maria C. Bossa<sup>2B,D</sup>,  
Anna Altieri<sup>2B,D</sup>, Cartesio Favalli<sup>1,2,G</sup>

<sup>1</sup> Department of Experimental Medicine and Biochemical Sciences, "Tor Vergata", University of Rome, Rome, Italy

<sup>2</sup> Clinical Microbiology Laboratories, Polyclinic of Tor Vergata, Rome, Italy

Source of support: Departmental sources

**Perché non utilizzare il sistema Uro-Quick nel pre-arricchimento routinario di tutti i campioni biologici liquidi?**

**Il prearricchimento su flaconcino ,condotto in parallelo con tecniche colturali tradizionali.**

- utilizzato con tempi di lettura pari a 235 min/6h, (tali da consentire la stima minima della carica batterica pari a <15 CFU/ml) e sempre in combinazione con lo studio del PAR.
- Per valutare e compensare nella coltura gli effetti di terapie antibiotiche impostate prima del prelievo del campione.



## Risultati su 546 campioni

Table 1. Comparative yields of clinically significant isolates of bacteria and yeast.

Specimens	No. of specimens (%)	No. of positive samples by routine culture	No. of negative samples by routine culture	No. of positive samples by URO-QUICK™	No. of negative samples by URO-QUICK™	p for URO-QUICK™ vs. routine method
ASB	106 (19.4%)	96	10	96	10	0.99
BAL	63 (11%)	58	5	58	5	0.99
Sputum	139 (25%)	134	5	136	3	0.28
Blood	47 (8%)	0	47	29	18	<0.000005
PLEF	105 (19%)	18	87	24	81	0.30
CSF	26 (4.7%)	0	26	2	24	0.49
PERF	41 (7.5%)	14	27	16	25	0.49
Other Fluid	19 (3.4%)	3	16	6	13	0.26
<b>Total</b>	<b>546</b>	<b>323</b>	<b>223</b>	<b>367</b>	<b>179</b>	<b>0.007</b>

ASB – endotracheal aspirates; BAL – bronchoalveolar lavage; PLEF – pleural fluid; PERF – peritoneal fluid; CSF – cerebrospinal fluid.

**+ 44 positivi**

## Isolati ottenuti solo con HB&L non da coltura (nei 44 campioni)

Table 2. Bacterial isolates uniquely obtained using URO-QUICK™.

Specimens	Total number of specimens (%)	Specimens uniquely positive by URO-QUICK™ (%)	Isolates (n)
ASB	106 (19.4%)	–	
BAL	63 (11%)	–	
Sputum	139 (25%)	2 (1.4%)	<i>Streptococcus pneumoniae</i> , <i>Pseudomonas aeruginosa</i>
Blood	47 (8%)	29 (61%)	<i>Sphingomonas paucimobilis</i> , <i>Serratia marcescens</i> , <i>Staphylococcus hominis</i> , <i>Staphylococcus epidermidis</i> SCV (n=5)*, <i>Staphylococcus aureus</i> (n=2), <i>P. aeruginosa</i> (n=2), <i>Oligella urealyticum</i> , <i>Micrococcus luteus</i> , <i>Moraxella lacunata</i> , <i>Enterococcus faecalis</i> (n=2), <i>Escherichia coli</i> (n=3), <i>Corynebacterium propinquum</i> , <i>Corynebacterium jeikeium</i> (n=2), <i>Clostridium tyrobutyricum</i> , <i>Candida albicans</i> (n=2), <i>Campylobacter jejunii</i> , <i>Acinetobacter baumannii</i>
PLEF	105 (19%)	6 (5.7%)	<i>S. aureus</i> , <i>Rhodotorula glutinis</i> , <i>Bacillus pumilus</i> , <i>Burkholderia cepacia</i> (n=2), <i>Burkholderia gladioli</i>
CSF	26 (4.7%)	2 (7.6%)	<i>E. coli</i> , <i>S. aureus</i>
PERF	41 (7.5%)	2 (7.8%)	<i>Gemella morbillorum</i> , <i>Enterococcus avium</i>
Other Fluids**	19 (3.4%)	3 (15.8%)	<i>Acinetobacter baumannii</i> - <i>Klebsiella pneumoniae</i> , <i>Pasteurella multocida</i> , <i>S. aureus</i>
Total	446	44 (8.0%)	

ASB – endotracheal aspirates; BAL – bronchoalveolar lavage; PLEF – pleural fluid; PERF – peritoneal fluid; CSF – cerebrospinal fluid.

\* Small colony variant (SCV); \*\* Other fluids (19 in total) included synovial fluid (n=5), ascitic fluid (n=9), drainage of infected central venous catheters (n=3), abdominal drainage (n=1), and cholecystic fluid (n=1).



## Interessanti le indicazioni fornite dal PAR sui liquidi

- PPV 100% , svelando anche terapie non dichiarate es in casodi meningite

16% dei campioni testati mostravano PAR non allineato alla terapia dichiarata:

- 67% Sensibilità
- 76% NPV

(16%) c'erano alcuni comuni denominatori:

- 1 Presenza di anti fungini, metronidazolo, o carbapenemi
- 2 La terapia era stata iniziata/terminata solo nelle 12 h precedenti il prelievo
- 3 Regime terapeutico inappropriato (dosaggi troppo bassi)
- 4 2 casi veri incongruenti in LCR in pazienti in terapia con Vancomicina



## Sorpasso sui tempi



Se campione monomicrobico (post Gram) si può allestire direttamente ID e AST

- Prearricchimento 235 min/6h + centrifugazione e recupero del pellet batterico su cui eseguire direttamente ID AST i tempi si contraggono di -18 ore!!!



## Se campione polimicrobico ci arrendiamo??

- La tecnologia degli ultimi anni ci è venuta in aiuto
- Arricchimento H&BL e sequenziamento misto

## 16S Direct Sequencing Identification from Mixed Clinical Samples

Isentio RIPSEQ is an online application that dramatically improves the bacteria identification process. A must for samples affected by Antibiotics and for Anaerobe Infections.

[Clinical use](#) [Product details](#)



View our demonstration video of RipSeq Mixed



Try a do-it-yourself demo of RipSeq Mixed right now!

### User meetings

**June 9th, Oslo, Norway:** RipSeq User Meeting, Thon Hotel Opera.

### Latest news

**Press release:** Isentio AS is expanding and opens office in Silicon Valley, USA

**Press release:** 1.5 million USD in new funding & new chairman of the board to Isentio AS

**Publication:** Simultaneous Sequencing Analysis of The 16S rRNA and rpoB Genes Using RipSeq Software to Identify Mycobacterium Species (by ARUP Labs & Isentio)

### New product!

**RipSeq Single:** [Single click batch analysis of DNA sequence files in seconds!](#)

The resulting product had a size of approximately 460 bp, covering the variable areas V1, V2, V3 and V4 of the 16S rRNA gene

## Analysis of Mixed Sequencing Chromatograms and Its Application in Direct 16S rRNA Gene Sequencing of Polymicrobial Samples<sup>▽</sup>

Øyvind Kommedal,<sup>1,2\*</sup> Bjarte Karlsen,<sup>3</sup> and Øystein Sæbø<sup>3</sup>

*Department of Microbiology and Immunology, Haukeland University Hospital, Bergen, Norway<sup>1</sup>; Section for Microbiology and Immunology, the Gade Institute, University of Bergen, Bergen, Norway<sup>2</sup>; and iSentio Ltd, Thormøhlensgate 51, Bergen, Norway<sup>3</sup>*

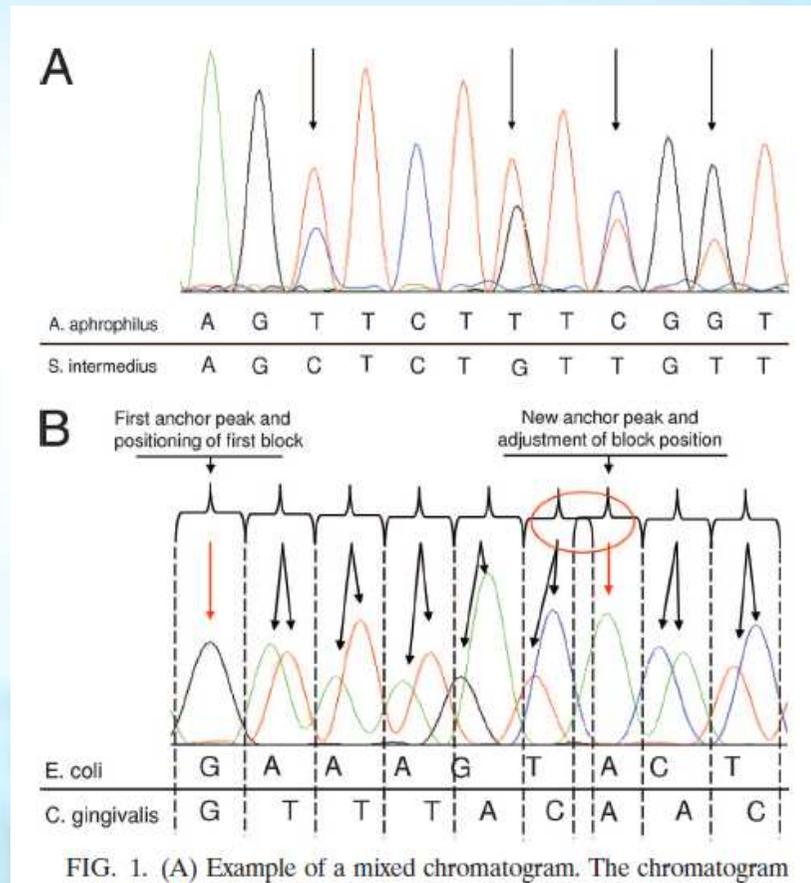
Received 2 February 2008/Returned for modification 4 April 2008/Accepted 23 August 2008

Investigation of clinical samples by direct 16S rRNA gene sequencing provides the possibility to detect nonviable bacteria and bacteria with special growth requirements. This approach has been particularly valuable for the diagnosis of patients who have received antibiotics prior to sample collection. In specimens containing more than one bacterium, direct sequencing gives mixed chromatograms that complicate further interpretation. We designed an algorithm able to analyze these ambiguous chromatograms and implemented it as a Web-based service. The algorithm contains both a new base-calling procedure and a new database search procedure. 16S rRNA gene sequencing was performed on polybacterial suspensions prepared in the laboratory. The computer program identified all bacteria correctly to the species level in 23 out of 23 samples containing two different bacteria. For samples containing three different bacteria, correct identification to the species level was achieved for three out of five and to the genus level for five out of five.

In our opinion, the most important feature of direct 16S rRNA gene sequencing is the possibility to analyze samples collected after the administration of antibiotics. Foci in inter-

The PCR reaction was optimized to reach a sensitivity of 1-10 genome copies per reaction tube. By spiking of human EDTA blood with different concentrations of bacteria, using the definition of a positive sample described above, the sensitivity of the assay was found to be 2000-4000 genome copies per ml of sample material.

unequally, permitting some to grow and not others.



Sequenza diretta da campione polimicrobico, gli elettroferogrammi misti conterranno due o più picchi sovrapposti nelle zone del 16S rDNA variabile e singoli nelle zone conservate.

Risultato il programma trascrive una

- sequenza normale + 1° base nella zona variabile;
  - sequenza normale + 2° base nella zona variabile; ecc
- Ovviamente combinando tutte le possibili variabili



## RIPSEQ MIXED

Mixed subscription ends: 30.05.2011

File:

**3025235 emoseq5-17-11-4-57 PM.ab1**

1. Basic settings 2. Advanced settings **3. Basecalling result** 4. Identification

Total length: 207, total number of bases: 567, 1:9 (4%), 2:71 (34%), 3:92 (44%), 4:35 (16%)

46				50				60											
G	A	A	A	A	A	A	C	C	C	C	C	C	C	G	A	A	G	C	A
	G	C	C	C	C	C	G	G	G	T	T	G	T	T	T	G	T	T	C
	T		G	T	T	T	T	T	T										G
																			C
																			A
555	567	579	591	603	615	627	641	653	663	675	687	698	710	721	732	744	756	768	780
																			792



Analyze now

<< Back to basic settings

<< Back to advanced settings



File:  
**emoc 326843-29-11-3-53 PM.ab1**

- 1. Basic settings
- 2. Advanced settings
- 3. Basecalling result
- 4. Identification**

X	Score	Hits	Genus	Species	Strain	Accession	Diff %	Link	BLAST	Bases
▶	100,0	344/344	Staphylococcus (F)	epidermidis						
▶	99,4	342/344	Proteus (F)	mirabilis						

**Petti, C. A., Bosshard, P. P., Brandt, M. E., Clarridge III, J. E., Feldblyum, T. V., Foxall, P., Furtado, M. R., Pace, N. & Procop, G. (2008a). *Interpretive criteria for identification of bacteria and fungi by DNA target sequencing; Approved guideline*. CLSI document MM18-A, 28. Wayne, Pennsylvania: Clinical and Laboratory Standards Institute.**

Comment for Save or Submit, if needed:

Signal cutoff (y-coordinate):

**Result saved OK**

# +ISENTIORIPSEQ

## Analysis results

Forward file: 1050488 emoseq5-17-11-5-35 PM.ab1

**Analysis date:** 19.05.2011 11:36:09  
**User:** Carla Fontana  
**Primer set 1:** 16S (4-531) primers - iSento  
**Solution set 1:** 16S rRNA Human Pathogens - iSento  
**User comment:** - none -

### Result

X	Score	Hits	Genus	Species	Strain	Accession
			Staphylococcus (R) aureus / Staphylococcus (R) capitis / Staphylococcus (R) caprae / Staphylococcus (R) cohnii / Staphylococcus (R) croceolyticus / Staphylococcus (R) epidermidis / Staphylococcus (R) haemolyticus / Staphylococcus (R) hominis / Staphylococcus (R) pettenkoferi / Staphylococcus (R) pseudolugdunensis / Staphylococcus (R) saccharolyticus			
1:	100,0	189/189				
X	100,0	189/189	Staphylococcus (R)	aureus	MSSA476	NC_002953
X	100,0	189/189	Staphylococcus (R)	capitis	ATCC 146	D83362
X	100,0	189/189	Staphylococcus (R)	caprae	ATCC 35538 T	AB009935
X	100,0	189/189	Staphylococcus (R)	cohnii	ATCC 29974 T	D83361
X	100,0	189/189	Staphylococcus (R)	croceolyticus	MCC 10046 T	AY953148
X	100,0	189/189	Staphylococcus (R)	epidermidis	MBRG 6.5	AJ514245
X	100,0	189/189	Staphylococcus (R)	haemolyticus	JCSC 1435	AP006716
X	100,0	189/189	Staphylococcus (R)	hominis	20021408	DQ056840
X	100,0	189/189	Staphylococcus (R)	pettenkoferi	A6664	DQ538520
X	100,0	189/189	Staphylococcus (R)	pseudolugdunensis	B006	EF053370
X	100,0	189/189	Staphylococcus (R)	saccharolyticus	ATCC 14953	L37602
	100,0	189/189	Staphylococcus (R)	capitis	ATCC 27840	L37599
	100,0	189/189	Staphylococcus (R)	capitis	GTC 727	AB233325
	100,0	189/189	Staphylococcus (R)	capitis	ATCC 49326 T	AB009937
	100,0	189/189	Staphylococcus (R)	caprae	DSM 20608	Y12593
	100,0	189/189	Staphylococcus (R)	epidermidis	MBRG 2.5	AJ508368
	100,0	189/189	Staphylococcus (R)	epidermidis	RP62A	CP000029
	100,0	189/189	Staphylococcus (R)	epidermidis	ATCC 12228	AE015929
	100,0	189/189	Staphylococcus (R)	hominis	GTC 1228 T	AB233326
	100,0	189/189	Staphylococcus (R)	hominis	ATCC 27844	L37601
	99,5	188/189	Staphylococcus (R)	auricularis	ATCC 33753	D83358

[Print simplified report](#)

[Print report with references](#)

Diff %	Link	BLAST	Bases

Staphylococcus (R) croceolyticus / Staphylococcus (R) epidermidis / Staphylo



## Un esempio

Nella notte: Paziente di 38 aa, arriva endocardite su protesi, febbrile e scompensato operato in urgenza rimossa e inviata la valvola , la vegetazione, tampone della vegetazione (in mezzo liquido)

Al mattino (7:30): Valvola e vegetazione seguono un percorso tradizionale

Il tampone viene inoculato in HBL  
Dopo 3 h già positivo si preleva una aliquota risulta coltura mista:  
Gram+/Gram-

a 6 h si procede poi con subcoltura

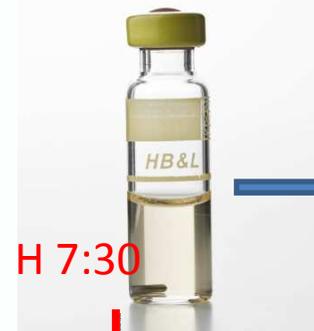
H 7:30

Subcoltura tradizionale con ID + AST a

H 14:30 sub coltura

H 8:00 del mattino successivo ID AST

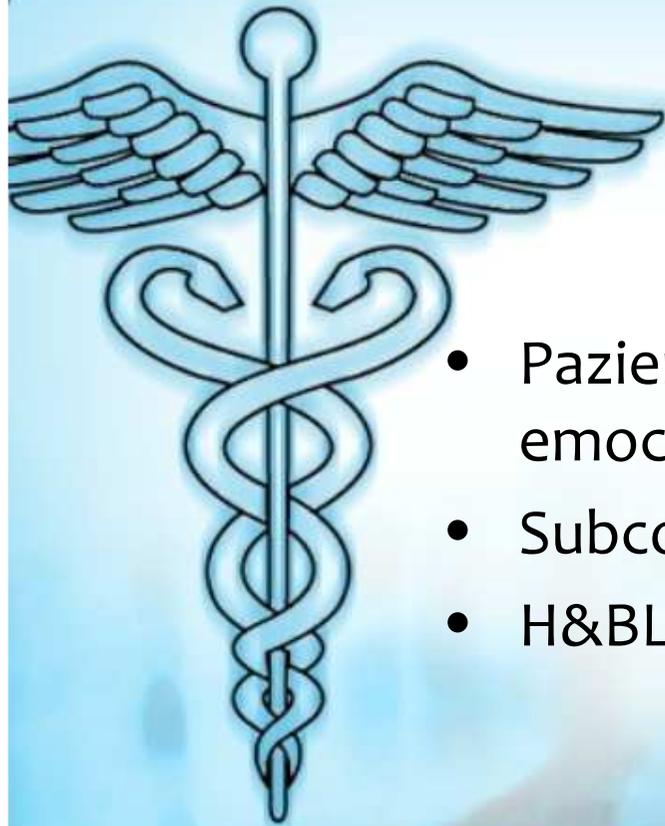
H 19:00 referto



H 7:30

11:00 Rip Seq  
(15 min, 1,5h,  
58min, 20 min)

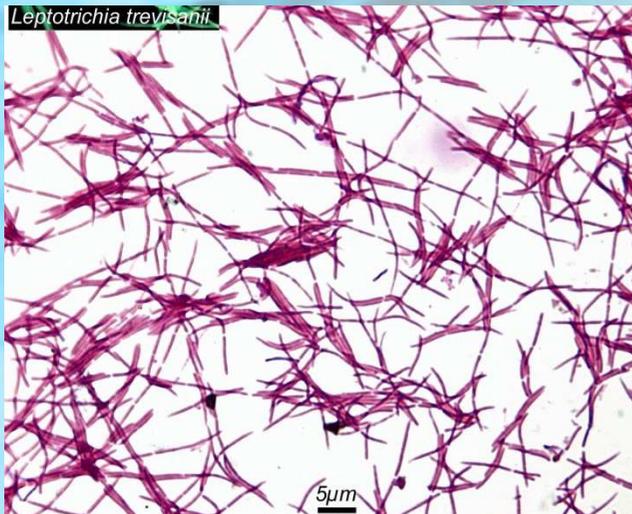
H 17:00 *S.aureus*,  
*Burkholderia pyrrocinia*



## Un altro esempio

- Paziente ematologica 41 aa con ripetute emocolture positive TD medio32.2 (bacilli Gram +)
- Subcolture ripetutamente negative
- H&BL in 6 h sequenziamento diretto e E-test

*Leptotrichia trevisanii*



La coltura tradizionale su piastra dopo oltre 48h di incubazione dalla subcoltura!

Germe che non avremmo potuto identificare per via tradizionale



## Germi più frequentemente recuperati con HB&L da soli ed in associazione con altri patogeni e persi in coltura tradizionale

CONS scv, Candida, GNNF	CVC
S.aureus, S.pneumoniae	Aspirati/Pus/Emocolture da endocarditi
Candida e C.glabrata	F.peritoneale/Pus/emocolture
Enterobatteri	Respiratori-Aspirati
Gram-neg NF	Pus-Aspirati Respiratori
C.Jk ed altri Corinebatteri	emocolture
B.cepacia	Aspirati
C.coli, C.jejunii	emocolture
Capnocytophaga/Actinomyces	emocolture
H.influenzae	emocoltura/respiratorio
L.monocytogenes, S.pneumoniae, N.meningitidis	LCR
S.bovis	emocolture
E.faecium	Pus



## Un altro esempio, in itinere

- Lesioni del piede diabetico
- IDSA 2012 ci indicano la stadiazione, la terapia e i campioni ideali
- Biopsie, frammenti di tessuto profondo
- T. da ferita, da effettuare solo in casi estremi (ma comunque utili se abbinato al Q-score)

2012 Infectious Diseases Society of America Clinical Practice Guideline for the Diagnosis and Treatment of Diabetic Foot Infections Benjamin A. Lipsky, et al



## Nostri primi risultati?!

- Arricchimento e-swab in H&BL
- Rip seq
- Coltura tradizionale

Coltura tradizionale	H&BL + Rip Seq	
<i>S.aureus</i>	<i>S. aureus</i>	<i>Prevotella disiens</i>
<i>A.baumannii</i>	<i>A.baumannii</i>	<i>Alcaligenes faecalis</i>
<i>S.epidermidis</i>	<i>S.epidermidis</i>	<i>E.faecalis</i>
<i>E.faecium</i>	<i>E.faecium</i>	<i>S.aureus</i>
<i>S.epidermidis</i>	<i>S.epidermidis</i>	<i>Brucella canis</i>
<i>E.raffinosis</i>	<i>E.raffinosis</i>	<i>C.parapsilosis</i>
<i>E.coli</i>	<i>E.coli</i>	<i>S.aureus</i>



er  
d

ere  
rica  
one  
odo  
mi



## Quale il messaggio? :

### il tempo è denaro

- Perché si riducono i tempi delle terapie empiriche
- Perché si corregge la terapia e si evitano ulteriori complicanze
- Perché si migliora outcome
- Perché si riducono i giorni di degenza
- Perché si evitano terapie inappropriate che pesano sulla epidemiologia d'ospedale



## Altro punto critico CRBSI

- L'uso dei cateteri venosi centrali (CVC) utilizzo sempre più diffuso.
- pro e cons: facilitano la gestione del paziente vs esposizione al rischio d'infezione

## Alcuni numeri



- Si stima che nel >78% ICU patients venga inserito come procedura di routine un CVC
- Circa 15 milioni CVCs sono mediamente inseriti per anno United States e le CRBSI sono ai primi posti fra le HAI circa 250.000 casi per anno di cui 80.000 in ICU, con costi 30,000 – 50,000 \$. \$2.3 billion annual cost to the U.S. healthcare system.
- incrementa il tasso di mortalità dal 12 al 25%
- Le CRBSI rappresentano da sole circa 87% delle BSI con un infection rate di 4.0-6.9 per 1000 giorni nelle ICU con il goal del CDC di scendere a <2 per 1000 gg catetere

PLoS ONE March 2012

Fletcher SJ: Central venous catheter related infection. *Anaesth Intensive Care*, 1999; 27: 425

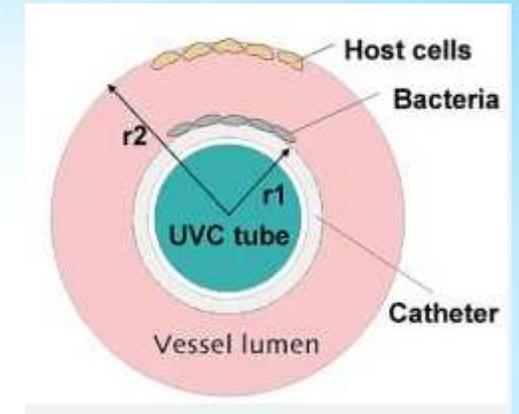
Mermel LA: Prevention of intravascular catheter-related infections. *Ann Intern Med* 2000; 132: 391–402

Wolf et al *Ann Hematol* 2005 DOI 10.1007/s00277-008-0509-5

CDC 2011

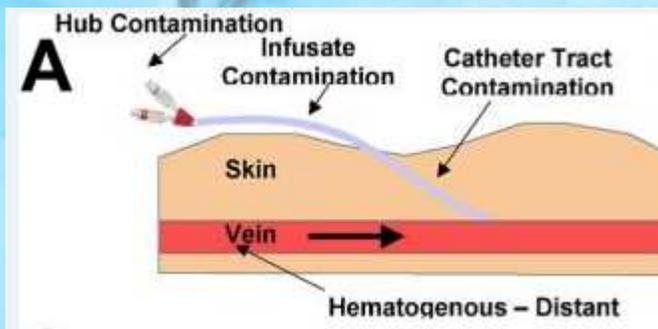


# Vie di infezione



## Ci sono 4 vie riconosciute di contaminazione:

- Migrazione dalla cute di organismi fino al sito d'inserzione e fin dentro la superficie interna con la colonizzazione della tip (più comune)
- contaminazione diretta del catetere o dell' hub via mani o con fluidi contaminati o altri devices
- il catetere può essere contaminato via ematogena da altro focus di infezione (meno comune)
- Contaminazione di infusi (rara)



CDC 2011

# Clinical Practice Guidelines for the Diagnosis and Management of Intravascular Catheter-Related Infection: 2009 Update by the Infectious Diseases Society of America

Leonard A. Mermel,<sup>1</sup> Michael Allon,<sup>2</sup> Emilio Bouza,<sup>9</sup> Donald E. Craven,<sup>3</sup> Patricia Flynn,<sup>4</sup> Naomi P. O'Grady,<sup>5</sup>  
Issam I. Raad,<sup>6</sup> Bart J. A. Rijnders,<sup>10</sup> Robert J. Sherertz,<sup>7</sup> and David K. Warren<sup>8</sup>

**Table 4. Commonly used clinical definitions of intravascular catheter-related infections.**

Infection	Definition
Catheter colonization	Significant growth of $\geq 1$ microorganism in a quantitative or semiquantitative culture of the catheter tip, subcutaneous catheter segment, or catheter hub
Phlebitis	Induration or erythema, warmth, and pain or tenderness along the tract of a catheterized or re-

Batteriemia/fungemia in un paziente con CVC e con almeno 1 emocoltura positiva con prelievo da VP accompagnata da manifestazioni cliniche quali febbre, brividi e /o ipotensione senza segni apparenti di infezione in altri distretti

Inoltre almeno uno dei seguenti parametri devono essere soddisfatti:

- Coltura CVC positiva con **>15CFU** per tip (se semiquantitativa) e **100CFU** se quantitativa l'isolato deve essere lo stesso da sangue e da coltura del CVC
- Oppure se è stato eseguito prelievo simultaneo VP e CV delta >3:1 nella stima CFU (cCVC vs VP) oppure delta nei TD d almeno 2h

or hypotension), and no apparent source for bloodstream infection (with the exception of the catheter). One of the following should be present: a positive result of semiquantitative ( $>15$  cfu per catheter segment) or quantitative ( $>10^2$  cfu per catheter segment) catheter culture, whereby the same organism (species) is isolated from a catheter segment and a peripheral blood culture; simultaneous quantitative cultures of blood with a ratio of  $>3:1$  cfu/mL of blood (catheter vs. peripheral blood); differential time to positivity (growth in a culture of blood obtained through a catheter hub is detected by an automated blood culture system at least 2 h earlier than a culture of simultaneously drawn peripheral blood of equal volume). Note that this definition differs from the definition of central line-associated bloodstream infection used for infection-control surveillance activities.

**NOTE.** Adapted in part from Pearson [18]. cfu, colony forming units.

<sup>a</sup> For surveillance purposes, patients with positive results of blood culture would be classified as having central line-associated bloodstream infection.



## La diagnosi

- Conservativa
- Non conservativa



## A rimozione avvenuta

Table 5.1 Microbiological techniques for diagnosis of CRBSI<sup>78</sup>

Technique	Description	Criteria for positivity	Sensitivity (%)	Specificity (%)
<b>Methods requiring device removal</b>				
Quantitative catheter tip culture	A distal tip segment of the removed device is flushed with broth or sonicated or vortexed in broth that is further incubated.	$\geq 100$ CFU	78-88	87-91
Semi-quantitative catheter tip culture	A 3–4cm distal tip segment of the removed device is rolled across an agar plate and incubated overnight. Unable to culture intraluminal organisms.	$>15$ CFU	81-89	85-87
Qualitative Catheter segment culture	Incubation of a segment of the removed device in broth media.	Any growth	79-96	72-78

Technique	Description	Criteria for positivity	Sensitivity (%)	Specificity (%)
<b>Methods not requiring device removal</b>				
Paired quantitative blood cultures	Paired blood cultures obtained through the device and from a separate venipuncture. Labour intensive.	Positive cultures from both sites and concentration of micro-organisms from the device 5 to 10-fold higher than from the peripheral venipuncture	74-84	98-100
Unpaired quantitative blood culture	Blood cultures obtained through the device.	≥100 CFU	80-93	83-89
Differential time to positivity	Concomitant conventional qualitative blood cultures obtained from the device and from a separate venipuncture continuously monitored until growth of microorganisms. Currently available with most automated blood culture systems. Hard to interpret when patient is taking antibiotics through the CVC.	Blood culture drawn through the device turns positive ≥120 min before those obtained from venipuncture	86-92	79-87
Acridine-orange leukocyte Cytospin on blood drawn through the device	Staining with acridine orange of a slide from 50µl blood and examined under ultraviolet light. Accuracy may be improved if performed on specimen obtained by endoluminal brushing.	Any microorganism within the cellular monolayer in a minimum of 100 high-power field	80-96	89-97
Unpaired qualitative blood culture	Blood cultures obtained through the device.	Any growth	84-98	83-89
Paired qualitative blood cultures	Paired blood cultures obtained through the device and from a separate venipuncture.	Any growth	51-65	78-95
Endoluminal brushing	Culture of sonicated and vortexed brush passed down the internal lumen to the device distal tip. May induce bacteraemia, arrhythmias, embolisation.	≥100 CFU	92-100	84-98
Culture of swabs of skin insertion site and of the hub	Semi quantitative cultures on agar plate.	Any growth	96-100	67-71

conservando





# La conservativa quali le procedure più usate ma quali i rischi

OPEN ACCESS Freely available online

PLoS one

CVC e da

## Catheter Related Bloodstream Infection (CR-BSI) in ICU Patients: Making the Decision to Remove or Not to Remove the Central Venous Catheter

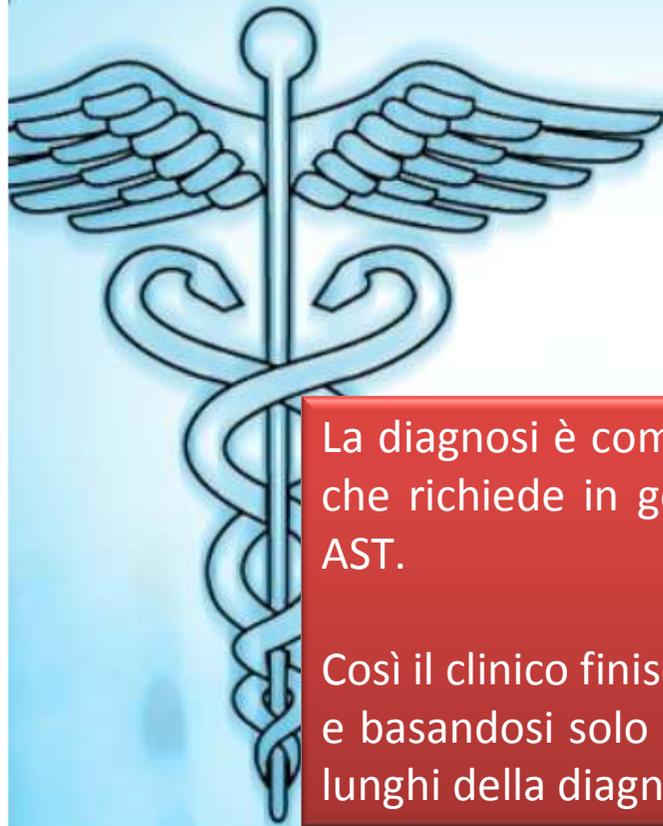
da CVC

Rodrigo Octávio Deliberato<sup>1\*</sup>, Alexandre R. Marra<sup>1</sup>, Thiago Domingos Corrêa<sup>1</sup>, Marinês Dalla Vale Martino<sup>2</sup>, Luci Correa<sup>3</sup>, Oscar Fernando Pavão dos Santos<sup>1</sup>, Michael B. Edmond<sup>4</sup>

<sup>1</sup> Critical Care Unit, Hospital Israelita Albert Einstein, Sao Paulo, Brazil, <sup>2</sup> Microbiology Laboratory Department, Hospital Israelita Albert Einstein, Sao Paulo, Brazil, <sup>3</sup> Infection Control Unit, Hospital Israelita Albert Einstein, Sao Paulo, Brazil, <sup>4</sup> Department of Internal Medicine, Virginia Commonwealth University School of Medicine, Richmond, Virginia, United States of America

### Conclusions

In case of CR-BSI it can be expected that prompt catheter removal will result in shorter duration of BSI and improved outcomes [24,25]. In our study there was a no statistically significant difference between the standard and conservative methods in-hospital mortality but there was a trend toward higher mortality rates among patients with CR-BSI diagnosed by the conservative method when the CVC was kept in place for more than 24 h. Further studies should be conducted to confirm this hypothesis.



## Un motivo del ritardo diagnostico

La diagnosi è comunemente basata sul risultato della emocoltura che richiede in genere almeno 48h + i tempi per esecuzione ID AST.

Così il clinico finisce per rimuovere il dispositivo al sospetto clinico e basandosi solo sui sintomi clinici non potendo attendere tempi lunghi della diagnosi conservativa

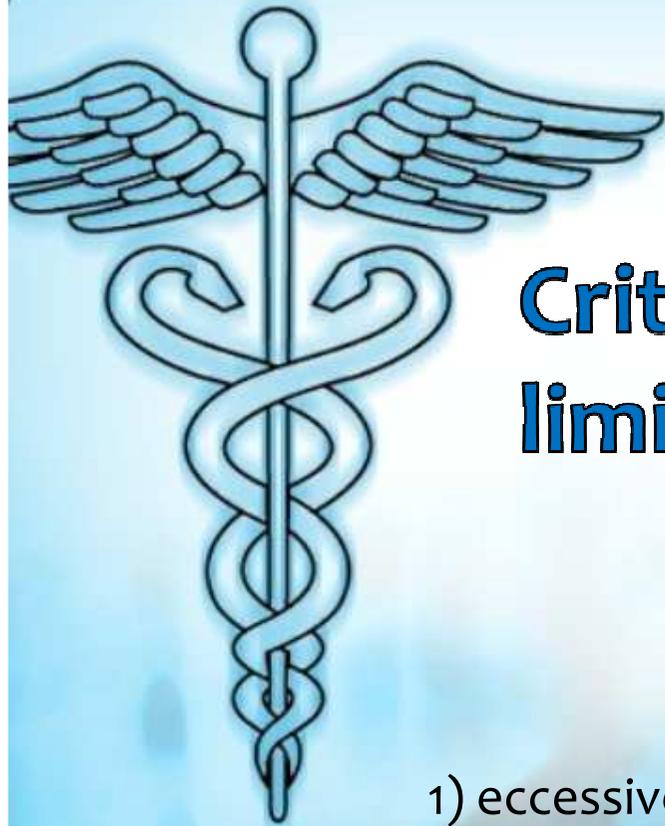


**Prudenza del clinico =  
eccessive rimozioni**

**Venous catheter microbiological monitoring. Necessity  
or a habit?**

**Piotr Smuszkiewicz<sup>1ADEF</sup>, Iwona Trojanowska<sup>1AEEG</sup>, Hanna Tomczak<sup>2ABCDG</sup>**

© Med Sci Monit, 2009; 15(2): SC5-8  
**PMID: 19179982**



## Criticita: eccessive rimozioni vs limiti della coltura

- 1) eccessive rimozioni, eccessivi invii di CVC al laboratorio
- 2) Molte evidenze scientifiche indicano come in numerosi casi di CRBSI pur a fronte di forti evidenze cliniche non sono poi sostenute dai dati microbiologici  
Così il numero di CRBSI accertate riportate in letteratura è compreso fra 1% - 40%



# IDSA 2009 : la coltura

Recommendation	Comments	recommandation	Reference(s)
Diagnosis: when and how should catheter cultures and blood cultures be done?			
Intravenous catheter cultures			
General			
1.	Catheter cultures should be performed when a catheter is removed for suspected CRBSI; catheter cultures should not be obtained routinely	A-II	[22, 26]
2.	Qualitative broth culture of catheter tips is not recommended	A-II	[22, 23]
3.	For central venous catheters (CVCs), the catheter tip should be cultured, rather than the subcutaneous segment	B-III	[20]
4.	For cultures of an anti-infective catheter tip, use specific inhibitors in the culture media	A-II	[31, 32]
5.	Growth of >15 cfu from a 5-cm segment of the catheter tip by semiquantitative (roll-plate) culture or growth of >10 <sup>2</sup> cfu from a catheter by quantitative (sonication) broth culture reflects catheter colonization	A-I	[22, 23, 27]
6.	When catheter infection is suspected and there is a catheter exit site exudate, swab the drainage to collect specimens for culture and Gram staining	B-III	[1, 33]



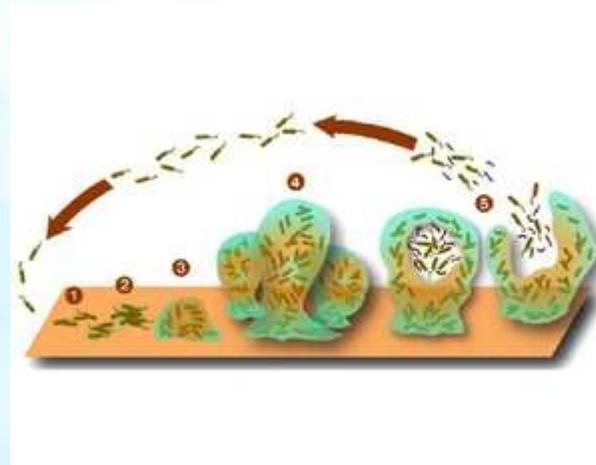
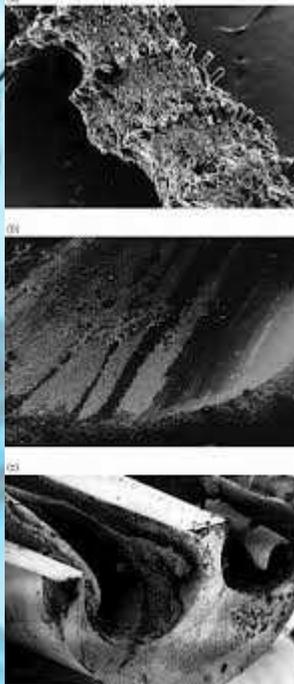
## Problematiche nella diagnosi: limiti della coltura

- Il problema principale risiede proprio nei limiti in termini di sensibilità della coltura:
- Maki's in particolare evidenzia per lo più i germi presenti sulla superficie esterna della tip, mentre per lo più il problema della colonizzazione e della formazione del biofilm è all'interno del lume
- I germi, per via delle terapie antibiotiche a cui sono esposti e soprattutto i biofilm produttori sono lenti nella crescita e i terreni di coltura solidi comunemente utilizzati non li supportano (differenti crescita di forme planktoniche e sessili)



# La biologia molecolare

- Forse ha un ruolo emergente





Research article

Open Access

## Use of cultivation-dependent and -independent techniques to assess contamination of central venous catheters: a pilot study

Mette KS Larsen<sup>1</sup>, Trine R Thomsen<sup>\*1,2</sup>, Claus Moser<sup>3</sup>, Niels Høiby<sup>3</sup> and Per H Nielsen<sup>1</sup>

### Maki a confronto tecniche di biologia molecolare

1. Passando attraverso la rimozione del biofilm interno ed esterno al CVC, estrazione da questo di DNA *16S rRNA gene amplification*

2. ovvero *Fluorescence in situ hybridization (FISH)* performed on fixed biofilm

## Biofilm esterno al CVC o del lume interno

Table 3: Clone library data

	Bacterial species	External	Internal	Mix
Alphaproteobacteria	<i>Aflpia broomeae</i>			3 (98%)
	<i>Bradyrhizobium japonicum</i>			5 (96–98%)
	<i>Bradyrhizobium</i> sp.			1 (96%)
Betaproteobacteria	<i>Acidovorax</i> sp.			3 (94–99%)
	<i>Alcaligenes</i> sp.			2 (96–98%)
Gammapro	<i>Pseudomonas</i> sp.		1 (97%)	
	<i>Serratia</i> sp.			2 (99%)
Deltaprotec Firmicutes				
Actinobacte				
Unknown	<i>Propionibacterium</i> species	2 (98%)		
	Uncultured bacterium clone 654931	2 (98–99%)		1 (99%)
	Uncultured organism clone MC060411			1 (95%)
	Total	62	62	35

Mix di batteri

However, many other bacteria belonging to the phyla *Proteobacteria*, *Firmicutes*, *Actinobacteria* and *Bacteroidetes* were also found, stressing that only a minor portion of the species present were found by cultivation

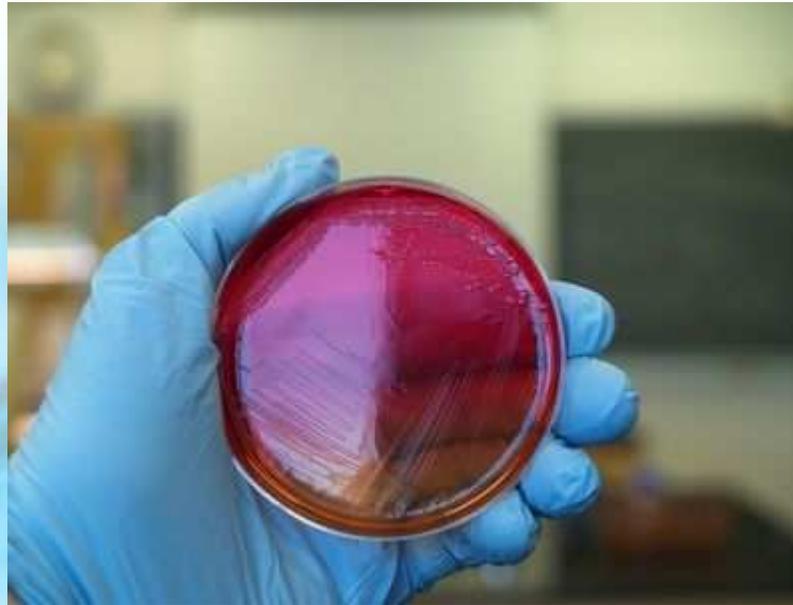
## Conclusion

The use of molecular techniques has the potential to substantially improve microbiological diagnosis in CVC-related infections when combined with existing methods based on cultivation and staining.

Blast results of the retrieved sequences from the three clone libraries from external, internal and mixed biofilm samples. Numbers refers to abundance of the different clones and the numbers in parentheses denote the percent ID, the clones have been identified with in the Blast search.



**Ma torniamo alla coltura**





**la nostra idea: i trials iniziali**



### Maki modificata

a) Semina diretta = rolling su plate

b) Semina Indiretta = vortex + in BHI, semina su piastra



semina diretta

semina indiretta

Maki



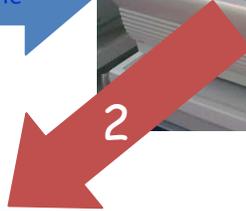
**PARTENZA 35% positivi**



# Nostra procedura



Rimozione del frammento, risospensione e 500 µl usati per inculcare H&BL  
Incubazione 6 h



Semina al termine delle 6 ore sul set di terreni solidi

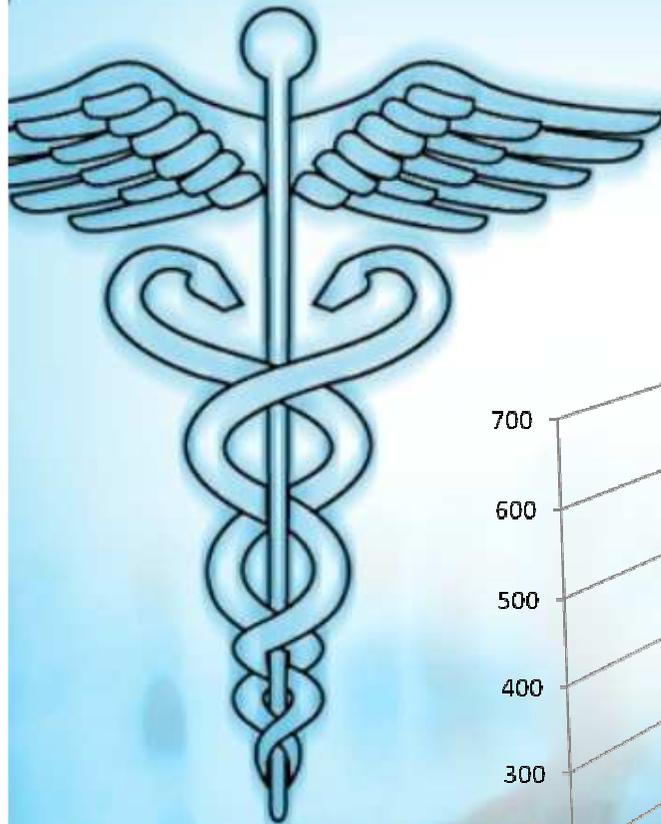
# Coltura tradizionale



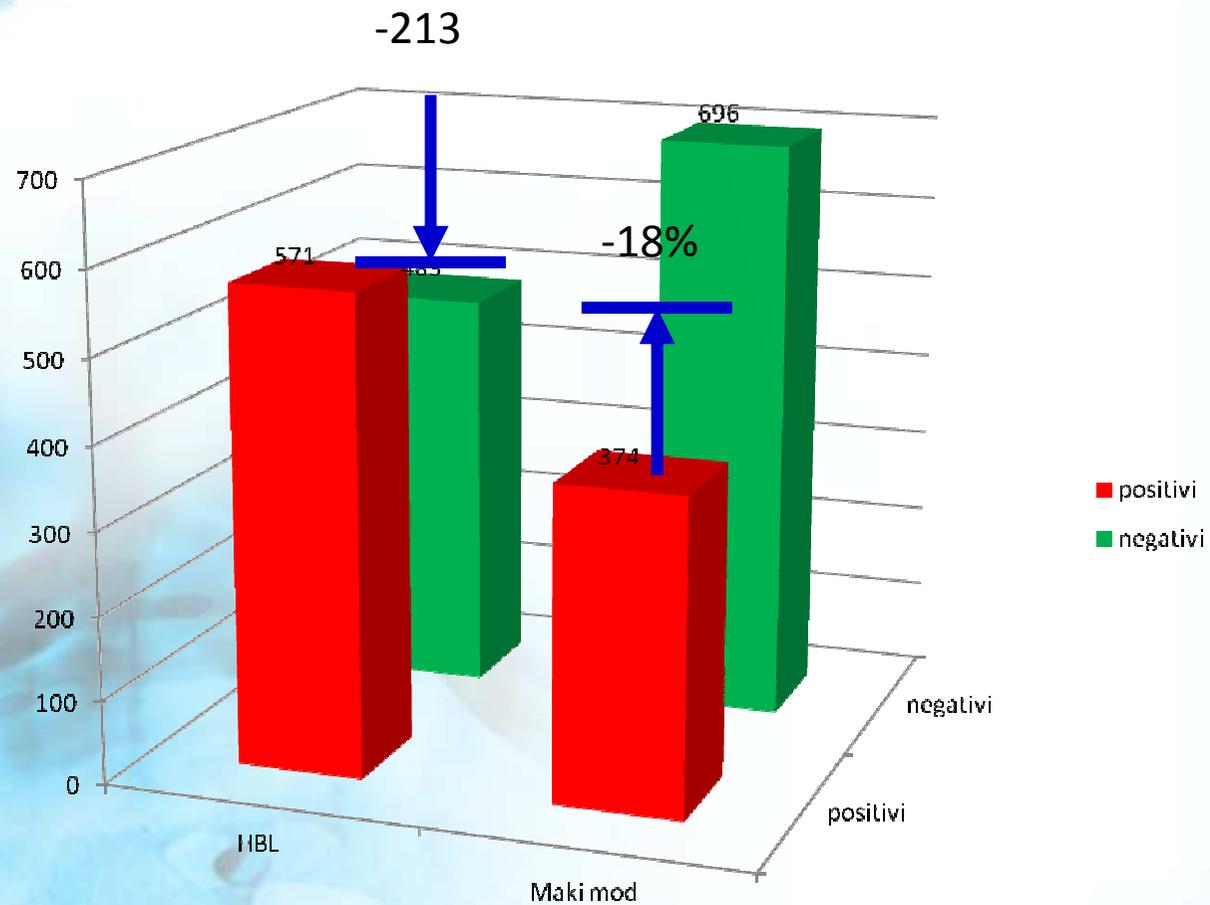


## La scelta

- I cateteri selezionati quelli a lunghi permanenza (tutti impregnati con chlorhexidine and silver sulfadiazine).
- esclusi i cateteri arteriosi
- Solo quelli dei pazienti per i quali disponevamo di emocolture nelle in un range di 48 h prima o dopo la rimozione



# 1070 cvc blood culture paired

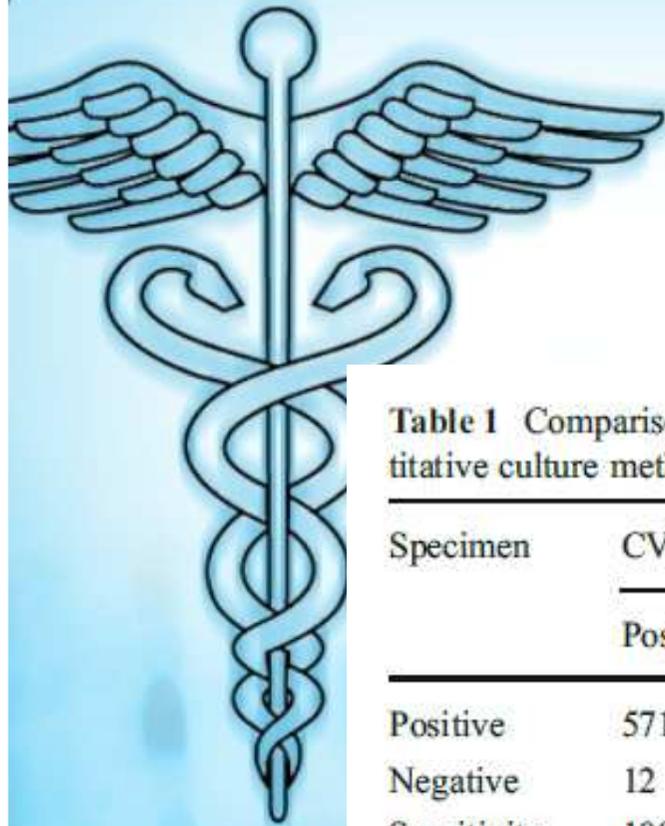


Eur J Clin Microbiol Infect Dis  
DOI 10.1007/s10096-012-1676-9

ARTICLE

## Improved diagnosis of central venous catheter-related bloodstream infections using the HB&L UROQUATTRO™ system

C. Fontana • M. Favaro • M. C. Bossa • S. Minelli •  
A. Altieri • M. Pelliccioni • F. Falcione • L. Di Traglia •  
O. Cicchetti • C. Favalli



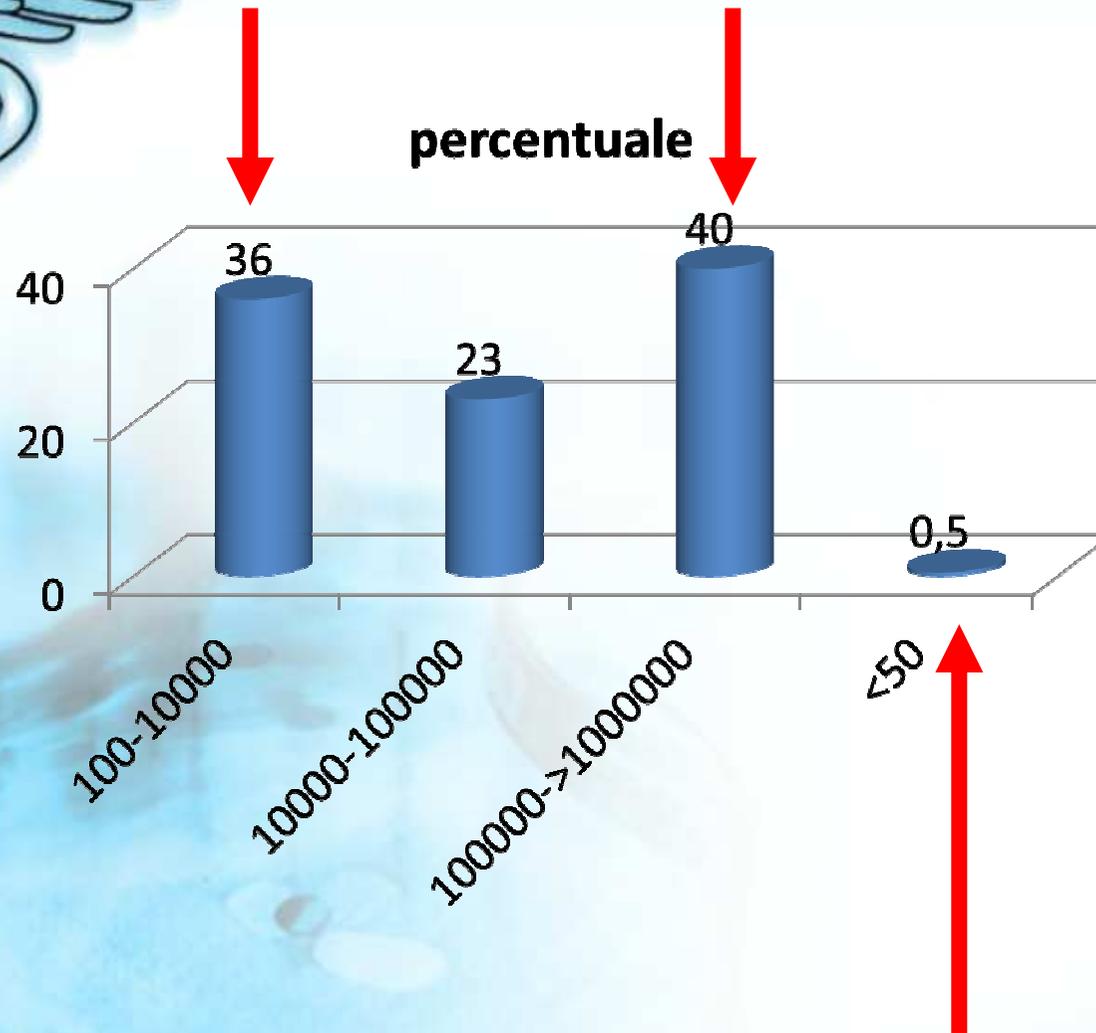
## HBL53.36 % vs Maki 34.95

**Table 1** Comparison of Maki's method with the CVC-HB&L™ quantitative culture method

Specimen	CVC-HB&L™ method		Maki's method	
	Positive	Negative	Positive	Negative
Positive	571	0	374	213
Negative	12	487	0	483
Sensitivity	100		63.71	
Specificity	97.59		100	
NPV	100		69.39	
PPV	97.94		100	
<i>p</i> -value	0.004			

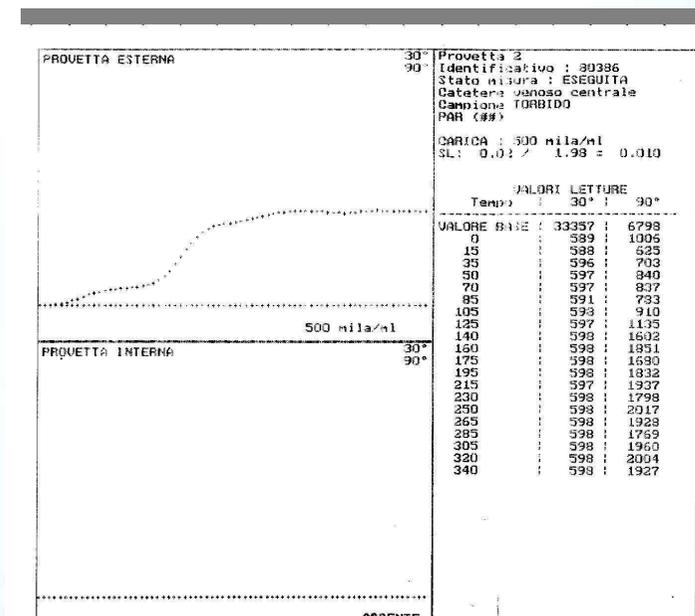
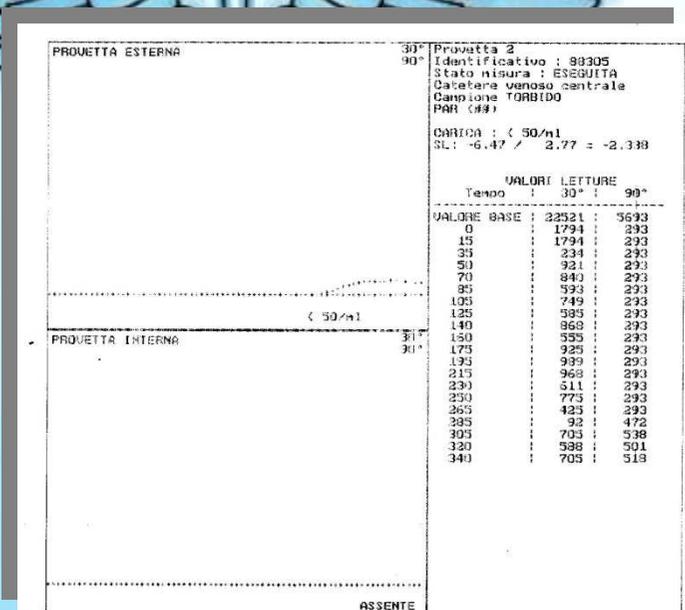
Of these patients, 571 patients were diagnosed with a CRBSI according to the current guidelines

Cut off più ragionevole  $\geq 100$  CFU



Le CFU stimate erano comprese in un range di  $1 \times 10^2$  to  $40 \times 10^6$  CFU/mL, con un mean value 602,266.667 SD di 151,294.9456.

## Interferenza da emazie (stime sovradimensionate)



12 campioni furono identificati come FP ma per le quali le emocolture erano **NEGATIVE**

E la subcoltura del brodino HB&L si mantenne negativa

Analizzando il campione si trattava di campioni con residui ematici e di altro materiale drenato dalla superficie interna del CVC tip che notoriamente interferiscono con la lettura HB&L UROQUATTRO™ instrument.

### ANALISI



NUOVI CAMPIONI

### OPZIONI



MODIFICA



INFORMAZIONI



ALTRO...



SETUP

Unità 1 - Pagina 1

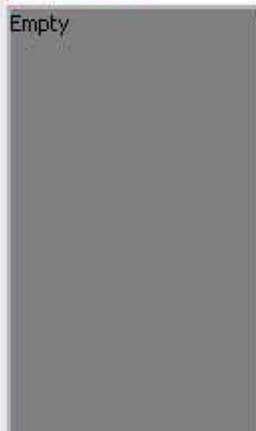


Table 2 Isolates from culture-positive CVCs

Species	No. of isolates on a total of 571 culture-positive CVCs using the CVC-HB&L™ method	No. of isolates on a total of 374 culture-positive CVCs using Maki's method
<i>Staphylococcus epidermidis</i> and other CONS	212+(10)	155 + (10)
<i>Staphylococcus aureus</i>	39	21
<i>Enterococcus faecalis</i>	78	35
<i>Enterococcus faecium</i>	36	22
<i>Klebsiella pneumoniae</i>	30+(3)	22 + (3)
<i>Klebsiella oxytoca</i>	3	2
<i>Proteus mirabilis</i>	13	7
<i>Escherichia coli</i>	20+(2)	18 + (2)
<i>Pseudomonas aeruginosa</i>	14	5
<i>Acinetobacter baumannii</i>	46+(4)	34+(4)
<i>Candida parapsilosis</i>	11+(1)	8+(1)
<i>Candida tropicalis</i>	8	4
<i>Candida glabrata</i>	3	2
<i>Candida albicans</i>	58+(1)	39+(1)
Total	592	395

The numbers in parentheses indicate cultures with two isolates per CVC

21 campioni erano polimicrobici co associazioni tipo Gram-positivi (CONS), Gram negativi (*Escherichia coli*, *Klebsiella pneumoniae*, or *Acinetobacter baumannii*) o con lieviti (*Candida albicans* *Candida parapsilosis*).

La conclusione.....

Soluzioni semplici, anche semplici cambiamenti delle abitudini possono rappresentare «la soluzione» del microbiologo per la gestione dei campioni critici del paziente critico

.. *l'immaginazione è più importante della*

*conoscenza*

*Albert Einstein*

